

# Cellular Neuro-physiology

## Synaptic integration of young neurons into the adult hippocampus

In the adult hippocampus new neurons are continuously generated throughout life by adult neural stem cells (Fig. 1). The newly generated young neurons show a number of distinct functional properties, including enhanced excitability, reduced GABAergic inhibition and enhanced synaptic plasticity (Lodge and Bischofberger 2019). Based on these cell biological findings it was concluded that the young neurons are hyperactive and hyperexcitable during learning behavior and memory processing. However, on the behavioral level adult neurogenesis improves learning by increasing the brains capability to distinguish between similar memory items, a process called “pattern separation”. During the last research period (2017–2020) we focussed on glutamatergic and GABAergic circuit analysis, to address this apparent paradox of “hyperactive cells” versus “improved pattern separation”.

Synaptic recruitment of adult-born young granule cells. We studied excitatory glutamatergic synaptic transmission from cortical perforant-path fibers onto newly generated young granule cells (GCs) up to 4 weeks post mitosis (wpm, Li *et al.* 2017). We found that the young neurons fire action potentials (APs) as early as 2 wpm in response to a small number of active glutamatergic synapses, due to a high synaptic gain. However, due to small dendritic trees and sparse connectivity, neighboring young neurons are activated by different distinct small subsets of afferent fibers with minimal overlap. As the neurons mature, the increase in synapse number is balanced by a gradual decrease in intrinsic excitability. This indicates that the enhanced excitability in young granule cells does not generate hyperexcitability, but instead compensates for sparse synaptic connectivity in developing young neurons. Using paired whole-cell recordings, we could show that AP firing in neighboring young cells is not unspecific, but rather dependent on small non-overlapping populations of afferent input fibers (Fig. 2). This is due to the sparse connectivity combined with the high synaptic gain, generating differential and highly specific spiking output in neighboring cells. Therefore, perforant-path fibers can recruit young neurons in a sparse and orthogonal manner, well suited to support sparse coding during hippocampal information processing.



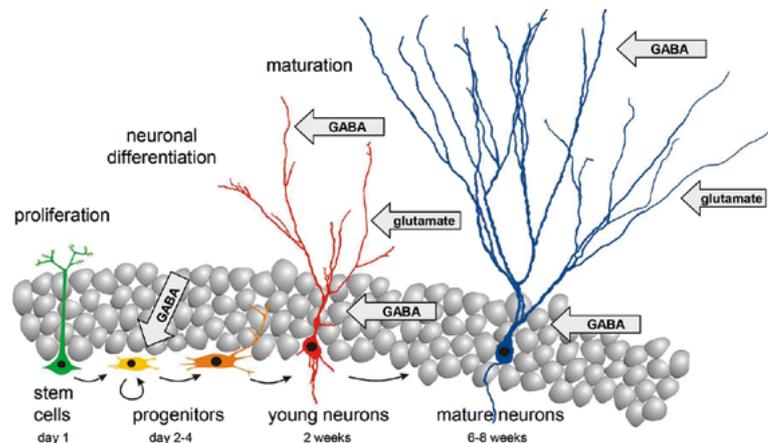
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**Fig. 1: Adult neurogenesis in the hippocampus.** Adult neural stem cells (green) are localized within the subgranular zone of the hippocampus and give rise to transient amplifying cells (yellow). They generate postmitotic neuroblasts which subsequently differentiate into young neurons (red). During the following ~6 weeks, they form new dendrites and several thousand new synaptic connections with glutamatergic neurons as well as with various types of soma- and dendrite-targeting GABAergic interneurons.

GABAergic interneurons. We further studied the function of soma- and dendrite-targeting GABAergic interneurons, identified by parvalbumin (PV) or somatostatin (SOM) expression, respectively (Schulz *et al.* 2018, 2019). Synapses onto young and mature GCs as well as CA1 pyramidal cells were investigated. Remarkably, in young GCs both, PV and SOM interneurons activated  $\alpha 5$ -subunit containing GABA receptors ( $\alpha 5$ -GABA<sub>R</sub>), showing a pronounced voltage-dependent outward rectification. As a consequence, the conductance was 4-fold larger at depolarized potentials close to AP threshold (40 mV) as compared to the resting potential. By contrast, in fully mature hippocampal neurons (GCs and CA1 pyramidal cells)  $\alpha 5$ -GABA<sub>R</sub>s are only involved in dendritic inhibition and fully excluded from PV interneuron synapses (Schulz *et al.* 2018, Lodge *et al.* BioRxiv, 2020). Although GABA is considered to be the major inhibitory transmitter in the adult brain, it depolarizes newly generated young neurons, due to a depolarized GABA<sub>AA</sub> reversal potential (-35 mV). Nevertheless, we could show, that GABA can still act as an inhibitory transmitter in young GCs, due to shunting inhibition (Lodge and Bischofberger, 2019). Although activation of a low number of synaptic  $\alpha 5$ -GABA<sub>AA</sub>R facilitates depolarization, strong activation of synaptic  $\alpha 5$ -GABA<sub>AA</sub>R can nevertheless inhibit AP firing in young neurons, similar to mature GCs, because of the voltage-dependent conductance profile of  $\alpha 5$ -GABA<sub>AA</sub>R, generating shunting inhibition.

In conclusion, our studies explain how sparse glutamatergic connectivity and fine-tuned excitation-inhibition balance via specific GABA<sub>AA</sub> receptors enable distinct activation of different young neurons. This will not only help to avoid hyperexcitability, but also supports sparse activity in the hippocampus to improve hippocampal pattern separation.

## Selected Publications

- Lodge M, Bischofberger J (2019) Synaptic properties of newly generated granule cells support sparse coding in the adult hippocampus. *Behav Brain Res.* 372:112036
- Schulz JM, Knoflach F, Hernandez MC, Bischofberger J (2019) Enhanced Dendritic Inhibition and Impaired NMDAR Activation in a Mouse Model of Down Syndrome. *J Neurosci.* 39:5210–5221.
- Schulz JM, Knoflach F, Hernandez MC, Bischofberger J (2018) Dendrite-targeting interneurons control synaptic NMDA-receptor activation via nonlinear  $\alpha 5$ -GABA<sub>AA</sub> receptors. *Nat Commun.* 9:3576. doi:10.1038/s41467-018-06004-8.
- Normann C, Frase S, Haug V, von Wolff G, Clark K, Münzer P, Dorner A, Scholliers J, Horn M, Vo Van T, Seifert G, Serchov T, Biber K, Nissen C, Klugbauer N, Bischofberger J (2018) Antidepressants Rescue Stress-Induced Disruption of Synaptic Plasticity via Serotonin Transporter-Independent Inhibition of L-Type Calcium Channels. *Biol Psychiatry.* 84:55–64.
- Li L, Sultan S, Heigele S, Schmidt-Salzmann C, Toni N, Bischofberger J (2017) Silent synapses generate sparse and orthogonal action potential firing in adult-born hippocampal granule cells. *Elife.* 6:e23612. doi:10.7554/eLife.23612.

